INTRODUCTION

Cholesterol is a crucial molecule for animal life: it modulates the integrity and fluidity of cell membranes, is involved in multiple signal transduction pathways, and is the source molecule for bile acids and steroid hormones.\(^1\)\(^,\)\(^2\) Excessive amounts of cholesterol can result in diseases such as atherosclerosis, caused by accumulation of cholesterol in the arteries.\(^3\) To prevent this accumulation, at least two pathways exist to remove cholesterol from the body via the feces—the hepatobiliary pathway, the better-known of the two, and transintestinal cholesterol excretion (TICE). In the hepatobiliary pathway, high-density lipoprotein (HDL) mediates the transport of cholesterol from peripheral tissues to the liver, where it is secreted via bile.\(^1\) In an attempt to increase HDL-mediated cholesterol excretion, various therapies have been tested that inhibit the cholesteryl ester transfer protein. Although they did raise HDL cholesterol, these inhibitors did not increase fecal sterol excretion in humans.\(^4\) This suggests that inducing HDL concentrations alone might be insufficient to stimulate cholesterol excretion, and that stimulation of the TICE pathway may prove to be a more effective strategy. This review provides a brief overview of the classic hepatobiliary pathway and discusses how stimulation of the TICE pathway may enhance cholesterol efflux in humans.

CHOLESTEROL EFFLUX BY TICE

Classically, cholesterol is excreted out of the body in a pathway that depends on HDL particles, which transport cholesterol from peripheral tissues to the liver. This process includes the assembly of HDL, the transport of cholesterol into the HDL particle mediated primarily by the ATP-binding cassette A1 (ABCA1) transporter, and hepatic HDL cholesterol uptake by the scavenger receptor class B type I (SR-B1). In the liver, cholesterol can be converted into bile acids that are secreted into the bile along with phospholipids and cholesterol.\(^5\) Normally, about 95% of the secreted bile acids and up to 80% of intestinal cholesterol are reabsorbed into the intestinal lumen by apical sodium bile acid transporter (ASBT) and Niemann-Pick C1-Like 1 (NPC1L1), respectively.\(^6\)\(^,\)\(^7\) The exclusive biliary origin of fecal neutral sterol was originally challenged almost a century ago by Sperry, who suggested that there must be another, nonbiliary source of fecal cholesterol.\(^8\) In 1967, Simmonds et al. perfused humans and reported that cholesterol was not only absorbed but also secreted by the intestine.\(^9\)

With the emergence of knockout mice, it became possible to study this intestinal cholesterol secretion we now call TICE in more detail. For instance, mice deficient for the phosphatidylincholine transporter ATP-binding cassette B4 (ABCB4) had an almost completely abolished biliary cholesterol flow, but their fecal cholesterol loss was hardly affected.\(^9\) Similar results were obtained with mice deficient for ATP-binding cassettes G5 and G8 (ABCG5 and ABCG8) that together constitute a sterol cotransporter heterodimer located in the apical membrane of the intestine. These mice still had more than half of the fecal sterol excretion of their wild-type littermates despite a 90% reduction in biliary cholesterol excretion.\(^10\) Similarly, Temel et al. found that mass fecal cholesterol loss was not affected in mice upon external biliary diversion.\(^12\) In all these
studies, TICE could be calculated as cholesterol excreted in the feces that was not from dietary or biliary origin. Van der Velde et al. determined TICE directly with isolated murine intestines, in which they observed cholesterol secretion predominantly located in the proximal part of the small intestine. Altogether, these studies validated that the intestine can indeed secrete cholesterol into the intestinal lumen.

In addition to mouse studies, TICE was also examined by Jakulj et al. using a stable isotope-based method adapted to humans. In their study of 15 mildly hypercholesterolemic but healthy males, approximately 100 mg of the 900-mg total daily amount of neutral sterols excreted by the feces was derived from the diet, whereas biliary cholesterol secretion and TICE contributed about 600 mg and 200 mg, respectively. Thus, almost one-fourth of the fecal cholesterol excretion in humans is TICE, at least in mildly hypercholesterolemic but healthy males.

Although the molecular mechanisms of TICE remain largely elusive, it is evident that four subsequent steps are involved: (1) cholesterol uptake by the basolateral membrane of the enterocyte; (2) translocation of cholesterol from the basolateral to the apical side of the enterocyte; (3) excretion over the apical membrane into the intestinal lumen; and (4) partial reabsorption of the secreted cholesterol by NPC1L1 followed by translocation (Figure 1). The roles of the first and third step have been addressed by various studies, and cholesterol absorption by NPC1L1 has been known for many years, as reviewed before. However, cholesterol translocation in the enterocyte has been largely neglected. Therefore, much of this review will discuss likely mediators of this process as well as methods to modulate the steps.

BASOLATERAL CHOLESTEROL UPTAKE AND APICAL CHOLESTEROL SECRETION

The specific source of cholesterol needed for TICE is not currently known. Mice deficient for both ABCA1 and SR-B1 lack HDL and have a defective hepatic HDL-cholesterol clearance. In these mice, TICE of radiolabeled cholesterol from HDL was similar to that of their wild-type littermates. These observations indicate that HDL is not essential as a cholesterol source or transport moiety for TICE. However, it cannot be excluded that HDL does exert this function under physiological circumstances and that a backup pathway with high affinity takes over HDL’s functions in the absence of this class of lipoproteins.

The cholesterol delivery or transport roles of very low-density lipoproteins (VLDL) or low-density lipoprotein (LDL) particles in TICE are not completely clear. Experiments with mice deficient for the LDL receptor (LDLR) do not show an effect on TICE, but interestingly, proprotein convertase subtilisin/kexin type 9 (PCSK9) knockout mice have an increased TICE while PCSK9 injections reduced TICE. This indicates that either the LDLR knockout mice upregulated an alternative pathway, or PCSK9 targets another receptor. In any case, these data suggest that VLDL/LDL cholesterol is a source for TICE.

It is evident that the ABCG5/ABCG8 sterol cotransporters, which are located in the apical membrane, play an important role in cholesterol excretion by the enterocyte into the intestinal lumen and are thus involved in TICE. Several studies have shown that TICE is strongly decreased in mice deficient for ABCG5 or ABCG8. The residual TICE in both knockout mouse models could be due to other apical cholesterol transporters, such as ABCB1.

PHARMACEUTICAL INTERVENTIONS THAT MODULATE BASOLATERAL OR APICAL CHOLESTEROL TRANSPORTERS

In accordance with the likely role of LDLR-mediated LDL uptake in TICE, pharmaceutical interventions (i.e., statins...
and PCSK9 inhibitors) that induce the LDLR protein have been shown to stimulate TICE in mice and ex vivo.\textsuperscript{18} Treatments that stimulate the expression of ABCG5/G8, such as pharmacological activation of the liver X receptor (LXR), have also been shown to stimulate TICE.\textsuperscript{15,20} Due to their lipogenic effect resulting in hepatic steatosis, the present LXR agonists have not yet been approved for use in humans.\textsuperscript{24} Intestinal LXR agonists that will not be absorbed beyond the enterocyte might prove to stimulate TICE in humans. Bile acids are ligands of the nuclear farnesoid X receptor (FXR), and they and other FXR ligands have been shown to stimulate TICE, likely by inducing the transcription of \textit{ABCG5} and \textit{ABCG8}.\textsuperscript{21,25} Finally, the anticholesterolemic drug ezetimibe has also been reported to stimulate fecal cholesterol excretion.\textsuperscript{14,23,26} Ezetimibe lowers intestinal cholesterol absorption via inhibition of the cholesterol influx transporter NPC1L1 located in the apical membrane. These studies suggest that part of the cholesterol excreted by the enterocyte is reabsorbed by enterocytes further down the intestinal tract.\textsuperscript{27}

**CHOLESTEROL ABSORPTION IN THE TICE PATHWAY**

Assuming that ezetimibe does exclusively inhibit NPC1L1, the magnitude of the drug’s effect on fecal cholesterol secretion provides information about the flux of cholesterol as it recycles through the enterocyte under normal conditions. In a study of mice and humans treated with ezetimibe, the amount of cholesterol reabsorbed by the enterocytes was at least twice the amount that was finally secreted via the feces.\textsuperscript{14} Thus, the amount of cholesterol secreted by TICE is apparently regulated at two sites: the excretion of cholesterol into the intestinal lumen counterbalanced by the reabsorption of cholesterol. This dual regulation of TICE could enable a more rapid regulation of the net TICE effect. Note that the estimated rate of cholesterol recycling in the enterocyte is probably too low because the enhanced secretion at the apical side of the enterocyte must be compensated by increased uptake of cholesterol at the basolateral side, which could become rate controlling. The high capacity of the intracellular cholesterol trafficking machinery may explain why changes in the rate of TICE have not been linked to changes in gene expression.

**TRANSLOCATION OF CHOLESTEROL THROUGH THE ENTEROCYTE**

Little is currently known about the molecular mechanism that enables cholesterol translocation between the basolateral and apical membranes of the enterocyte. When the transport follows a receptor-mediated pathway, cholesterol will be released from late endosomes/lysosomes via the coordinated action of Niemann-Pick type C1 and C2 proteins.\textsuperscript{28} However, it is unknown how cholesterol is subsequently released from the lysosomal membrane and which proteins/vesicles are involved in the intracellular transport. Lipid-transfer proteins (LTPs) have been shown to play a role in these processes, as they have the ability to bind lipids within a hydrophobic fold to mediate their transport.\textsuperscript{29} Among the LTPs, the likely sterol transporters are GRAM domain-containing protein 1 (GRAMD1) isoforms, oxysterol-binding protein (OSBP), OSBP-related protein 1 long form (ORP1L), steriodogenic acute regulatory protein (StAR), STAR-related lipid transfer (START) domain containing 3 (STARD3), and testis-expressed protein 2 (TEX2).\textsuperscript{30} It is clear from publicly accessible databases containing the immunohistochemical staining patterns of 76 human cell types, or the microarray profiles of 79 human or 61 mouse tissues, that GRAMD1 isoforms, OSBP, ORP1L, STARD3, and TEX2 might be involved in TICE since their protein and/or mRNA is found in the small intestine of mice and humans (Table 1).\textsuperscript{31-33}

<table>
<thead>
<tr>
<th>LIPID-TRANSFER PROTEIN</th>
<th>MOUSE</th>
<th>HUMAN</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>MRNA</td>
<td>MRNA</td>
</tr>
<tr>
<td>GRAMD1a</td>
<td>92</td>
<td>6</td>
</tr>
<tr>
<td>GRAMD1b</td>
<td>68/89*</td>
<td>88</td>
</tr>
<tr>
<td>GRAMD1c</td>
<td>514</td>
<td>8</td>
</tr>
<tr>
<td>OSBP</td>
<td>3493</td>
<td>14/146</td>
</tr>
<tr>
<td>ORP1L</td>
<td>30/94</td>
<td>36/176</td>
</tr>
<tr>
<td>StAR</td>
<td>5/5/11</td>
<td>5</td>
</tr>
<tr>
<td>STARD3</td>
<td>228</td>
<td>7</td>
</tr>
<tr>
<td>TEX2</td>
<td>5/595</td>
<td>36</td>
</tr>
</tbody>
</table>

* More than one value is due to multiple primers for the gene.

Table 1. Expression of lipid-transfer proteins in the small intestine of mice and humans. The higher the expression, the higher the value for messenger RNA expression.\textsuperscript{31,32} ND: not detected; GRAMD1: GRAM domain-containing protein 1; OSBP: oxysterol-binding protein; ORP1L: OSBP-related protein 1 long form; StAR: steriodogenic acute regulatory protein; STARD3: STAR-related lipid transfer (START) domain containing 3; TEX2: testis-expressed protein 2.
LIPID-TRANSFER PROTEINS IMPLICATED IN TICE

GRAMD1 Proteins

Multiple functions of the endoplasmic reticulum (ER) depend on its contact with other organelles via membrane contact sites (MCSs). Within these MCSs, the distance between the organelles is less than 30 nm, allowing localization of lipids and proteins involved in organelle functioning. This localization requires protein or protein complex tethers in the MCS that bind the two membranes. GRAMD1a is part of a tether between the ER and the plasma membrane (PM). The GRAMD1a ER-PM contact site contains a domain that has been found to be involved in the intermembrane transfer of sterols in yeast. The mammalian GRAMD1 isoforms also transfer sterols, but it is unclear if they are involved in TICE. Because of its high intestinal expression in both mice and humans, the GRAMD1b isoform could be a good candidate for involvement in TICE sterol flux (Table 1). Unfortunately, no pharmaceutical compounds are known to increase GRAMD1 activity. However, any future GRAMD1 activator designed for inducing TICE should be intestinal specific since GRAMD1 proteins are ubiquitously expressed, and inducing their activity outside the intestinal tract might have deleterious effects.

OSBP and ORP1L

OSBP is a cytosolic protein that can bind cholesterol, oxysterols, and phospholipids. It is best known for its cholesterol-transporting properties between the ER and the Golgi apparatus; it localizes to the apposed membranes of these two organelles and exchanges cholesterol for phosphatidylinositol 4-phosphate (PI4P), subsequently enriching the Golgi membrane. To form the ER-Golgi tether, OSBP must interact with the vesicle-associated membrane protein-associated protein (VAP) anchored in the ER. OSBP and OSBP-related proteins (ORPs) such as ORP1L share a C-terminal oxysterols-binding domain and an N-terminal pleckstrin-homology domain important for the membrane interaction. Interestingly, a loss-of-function of ORP1L in humans has been associated with low HDL-mediated cholesterol efflux, while ORP1L overexpression in macrophages reduced cholesterol flux to HDL in LDLR-deficient mice. Whether and how enterocyte OSBP and/or ORP proteins are critically involved in TICE needs to be investigated. A few compounds have been reported to inhibit OSBP and ORP proteins, and it would be of interest to test whether they affect TICE.

START Proteins: STARD3

STARD3 is part of a family of proteins that contains a START domain, named after the StAR-related lipid transfer. StAR, alias STARD1, regulates cholesterol transfer within mitochondria, a rate-limiting step in steroid hormones synthesis. Currently, 15 members of the START family have been identified in mammals. STARD3 is anchored at membranes of late endosomes and forms a tether with the ER-anchored VAP to enable cholesterol transport within the MCS. As such, STARD3 mediates cholesterol transport from the ER to late endosomes to maintain low cholesterol concentrations in the ER. Although members of the START family have not been shown to play a role in TICE, they have been implicated in cholesterol and lipoprotein metabolism. In hepatocytes, for example, phosphatidylcholine transfer protein, alias STARD2, transfers phospholipids to ABCB4 to facilitate their excretion via bile. Moreover, it is also suggested that cholesterol delivery to the hepatic ABCG5/ABCG8 transporter may depend on a yet unidentified START protein. Most of the cholesterol excreted over the apical membrane of the enterocyte is also transported by ABCG5/ABCG8, thus a role for START proteins in TICE is certainly not beyond imagination, with STARD3 as a good candidate.

TEX2

A fourth type of lipid-transfer protein contains a synaptotagmin-like mitochondrial-lipid-binding (SMP) domain. A well-known member of this family is cholesteryl ester transfer protein, which facilitates transfer of cholesteryl esters and other lipid species between the different circulating lipoproteins. TEX2, another member of the SMP family, might be a cytosolic sterol transporter, but it is mainly recognized as a transporter of ceramides. Hardly anything is known about the function of TEX2, especially in mammals. In yeast, TEX2 is part of a tether between the membranes of the Golgi and ER and between the nuclear envelop and vacuoles. As is evident for this obscure protein, more work is required to decipher its role in mammals before determining its role in TICE.

CONCLUSION

TICE is a nonbiliary route of excreting excess cholesterol out of the body via feces. Proteins involved in cholesterol uptake and efflux from enterocytes have been identified. It is probable that LDL and/or VLDL deliver cholesterol to the basolateral membrane of the enterocyte, while ABCG5/ABCG8 in the apical membrane transfer cholesterol into the intestinal lumen. Existing pharmaceutical compounds that modulate LDL uptake or ABCG5/ABCG8 will very likely affect TICE in humans. However, the machinery involved in intracellular trafficking of cholesterol remains enigmatic. Lipid-transfer proteins such as GRAMD1 isoforms, OSBP, ORP1L, STARD3, and TEX2 might be involved in this process. More dedicated research is eagerly awaited to test whether these lipid-transfer proteins indeed mediate TICE.
Conflict of Interest Disclosure:
The authors have completed and submitted the Methodist DeBakey Cardiovascular Journal Conflict of Interest Statement and none were reported.

Keywords:
transintestinal cholesterol excretion, TICE, NPC1L1, lipid-transfer proteins, GRAMD1, OSBP, ORP1L, STARD3, TEX2

REFERENCES


